

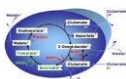
# Mitochondrial respiration medium - MiR06

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**Section**



**Page**

1. Mitochondrial Respiration Medium MiR06 .....	1
2. References .....	4

## 1. Mitochondrial respiration medium MiR06

**MiR06 = MiR05 + Catalase**; total volume = 1 litre (see Gnaiger et al 2000).

Compounds in MiR05/MiR06	Final conc.	FW	Addition to 1 litre final volume	Source; storage temp.
EGTA	0.5 mM	380.4	0.190 g	Sigma E 4378 (25 g); RT
MgCl <sub>2</sub> ·6 H <sub>2</sub> O	3 mM	203.3	0.610 g	Scharlau MA 0036 (250 g); RT
Lactobionic acid	60 mM	358.3 free acid	120 ml of 0.5 M <a href="#">K-lactobionate stock*</a>	Aldrich 153516 (100 g); RT
Taurine	20 mM	125.1	2.502 g	Sigma T 0625 (25 g); RT
KH <sub>2</sub> PO <sub>4</sub>	10 mM	136.1	1.361 g	Merck 104873 (1 kg); RT
HEPES	20 mM	238.3	4.77 g	Sigma H 7523 (250 g); RT
D-Sucrose	110 mM	342.3	37.65 g	Roth 4621.1 or Sigma 84097** (1 kg); RT
BSA, essentially fatty acid free	1 g/l		1 g	Sigma A 6003 fraction V (25 g); 4 °C

Weigh given amounts of the listed compounds (except BSA and lactobionic acid) into a 1000 ml glass beaker, disrupt big lumps mechanically, add 800 ml H<sub>2</sub>O and dissolve with magnetic stirring at 30 °C. Add 120 ml of K-lactobionate stock solution; adjust the pH to 7.1 (5 M KOH; Sigma P 1767; 1 kg) at 30 °C. Dissolve BSA separately in a subsample of the solution (recommended to prevent foam building) and transfer back to the final solution, while stirring continuously but gently. Adjust with H<sub>2</sub>O to a final volume of 1 litre. Check pH again and adjust if necessary. Store frozen at -20 °C in plastic vials.

**\* Preparation of K-lactobionate stock solution:**

Dissolve 35.83 g lactobionic acid in 100 ml H<sub>2</sub>O, adjust pH to 7.0 with KOH, adjust volume to 200 ml with H<sub>2</sub>O.

**\*\* Sucrose:**

Contamination with glucose may be critical. Invert sugar (1:1 mixture of glucose and fructose) is specified for Roth 4621 (<0.04%), but not for Sigma 84097. Invert sugar content is specified for Sigma 84100 (<0.05%), however the overall specified purity for this product (≥99%) is lower than for Sigma 8407 and Roth 4621.1 (both products ≥99.5%).

**MiR06** contains the following final concentrations:

Ca <sup>2+</sup> free	0.0 μM
Mg <sup>2+</sup> free	2.1 mM
K <sup>+</sup>	90 mM
Na <sup>+</sup>	0
EGTA free	0.46 mM
Osmolarity	330 mOsm
Ionic strength	95 mM

The ionic strength increases with the addition of substrates and adenylates, particularly in SUIT protocols with multiple substrate titrations.

**EGTA**

A general chelator for heavy metals, with high affinity for Ca<sup>2+</sup> but low affinity for Mg<sup>2+</sup>.

**Mg<sup>2+</sup>**

Activation by ATP due to ATPase activity is Mg<sup>2+</sup> dependent. The high quality of mitochondrial preparations cannot be tested in the absence of Mg<sup>2+</sup>. Physiological Mg<sup>2+</sup> concentration is in the range of 1-3 mM. Several enzyme systems depend on free Mg<sup>2+</sup>.

**P<sub>i</sub>**

The *K*'<sub>m</sub> is up to 1 mM in the ADP-activated OXPHOS state in heart mitochondria with glutamate/malate; 90% of maximum flux are reached at 10 mM. Is flux *measurably* higher at 15 mM?

**K-lactobionate**

The intracellular K<sup>+</sup> concentration is high (>100 mM), adding significantly to the ionic strength. In many previous studies of isolated mitochondria, KCl was used for this reason, but a high Cl<sup>-</sup> concentration is

unphysiological and inhibitory on mitochondrial creatine kinase (and possibly on other enzymes in the intermembrane space). K-MES or K-methanesulfonate have been used successfully. Lactobionate is well established in (extracellular) organ preservation solutions (University of Wisconsin solution).

### Taurine

Biological membrane stabilizer and ROS scavenger. 20 mM intracellular concentration in heart.

### Histidine

20 mM histidine was added in MiR04 (as in MiP02), as an imidazol-based buffer, with temperature dependence of pK identical to that of water ( $\alpha$ -stat pH buffer). No effect was observed when adding histidine to MITOMED1 with unpermeabilized endothelial cells. Omitted in MiR05 and MiR06 owing to increased autoxidation of TMPD and ascorbate in MiR04.

### HEPES

Well established buffer with pK close to 7.

### Sucrose

Impermeant and oxygen radical scavenger.

### BSA

Bovine serum albumine is a membrane stabilizer, oxygen radical scavenger, and binds  $\text{Ca}^{2+}$  and free fatty acids, hence the rather expensive essentially free fatty acid free BSA is required.

### Glutathione

No effect was observed with glutathione added to MITOMED1 with unpermeabilized or permeabilized endothelial cells (tEC); but background oxygen flux is significantly higher. This complication is avoided by not adding glutathione to the respiration medium.

### 98-03-30/04-02

Autoxidation of glutathione in MiR04 (with histidine) was variable between experiments, with  $a^{\circ'}$  of  $-1.7$  (range  $-2.1$  to  $0.3$ ,  $N=6$ ), and  $b^{\circ'}$  of  $0.053$ ,  $0.065$  and  $0.11$  in three oxygraphs (two chambers each).

### Catalase

is an antioxidant enzyme at high intracellular activity, and improves the antioxidant quality of physiological respiration medium beyond MiR05. In the presence of high catalase activity, hydrogen peroxide,  $\text{H}_2\text{O}_2$ , is titrated into the O2k-chamber to increase oxygen levels by up to  $200 \mu\text{mol/l}$ , e.g. from  $200 \mu\text{M}$  to  $350 \mu\text{M}$ . MiR06 is stored like MiR05, or can be prepared by adding catalase stock solution (dissolved in MiR05) directly into the closed O2k-chamber filled with MiR05 at the start of an experiment.

Compound	Final conc.	FW	Stock solution	Addition to 2 ml final volume	Source and product code
Catalase lyophilized powder, 2,000-5,000 units/mg protein* $\text{H}_2\text{O}_2$	280 $\mu\text{g/ml}^*$	34.01	112000 $\mu\text{g/ml}^*$ dissolve in MiR05 200 mM in $\text{H}_2\text{O}$ , adjust to	5 $\mu\text{l}$	Sigma C9322  Sigma Aldrich 516813 17.6

		pH 6	M, 50% w/w
	* Units of enzymatic activity (u) in $\mu\text{mol}/\text{min}$ ; assay used by Sigma Aldrich: 'One unit will decompose 1.0 $\mu\text{mole}$ of $\text{H}_2\text{O}_2$ per min at pH 7.0 at 25 °C, while the $\text{H}_2\text{O}_2$ concentration falls from 10.3 to 9.2 mM, measured by the rate of decrease of $A_{240}$ .'		
<b>H<sub>2</sub>O<sub>2</sub></b>	Small volumes ( $\mu\text{l}$ ) of $\text{H}_2\text{O}_2$ are injected into the O2k-chamber filled with MiR06, to increase oxygen levels. A typical $\text{H}_2\text{O}_2$ stock concentration in the syringe is approximately 200 mM. Manual injection: Fill a 10 $\mu\text{l}$ syringe with the $\text{H}_2\text{O}_2$ solution, inject 2-3 $\mu\text{l}$ , observe the oxygen level displayed by DatLab and inject stepwise ( $\Delta\text{CO}_2 \leq 200 \mu\text{mol}/\text{l}$ ) further $\text{H}_2\text{O}_2$ until the targeted oxygen level is reached. Alternatively, the "Oxystat" setup of the TIP2k may be used to reach a defined oxygen level and then maintain the oxygen concentration automatically between set limits in the oxystat-titration mode.		
<b>Creatine</b>	<b>MiR06Cr:</b> MiR06Cr = MiR06 plus 20 mM creatine. MiR05Cr or MiR06Cr has to be prepared fresh every day from MiR05 or MiR06. Do not freeze to avoid precipitation.		

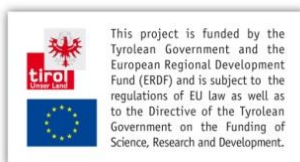
**Oxygen solubility factor** in MiR05 [1] at 30 °C and 37 °C is 0.92 [3]. The same solubility is valid for MiR06 and MiR06Cr.

## 2. References

- Gnaiger E, Kuznetsov AV, Schneeberger S, Seiler R, Brandacher G, Steurer W, Margreiter R (2000) Mitochondria in the cold. In: Life in the Cold. (Heldmaier G, Klingenspor M, eds) Springer, Heidelberg, Berlin, New York, pp. 431-42.
- Rasmussen HN, Rasmussen UF (2003) Oxygen solubilities of media used in electrochemical respiration measurements. *Analyt Biochem* 319: 105-13. *The manuscript (Gnaiger et al 2000) was sent to Dr. H. Rasmussen, and we appreciate that the oxygen solubility of MiR05 was then determined and published (Rasmussen 2003). Surprisingly, no reference was made in ref. (Rasmussen et al 2003) to the original publication on MiR05 (MiPNet08.05).*

[MiPNet03.02](#)

Selected media and chemicals for respirometry with mitochondria and permeabilized cells.



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[http://www.bioblast.at/index.php/MitoCom\\_O2k-Fluorometer](http://www.bioblast.at/index.php/MitoCom_O2k-Fluorometer)

### Author contributions

Gnaiger E with collaboration of Kuznetsov AV was responsible for the development and testing of MiR05 (Gnaiger et al 2000). All authors contributed to various details in the development of MiR06.

### Supplementary material:

[http://wiki.oroboros.at/index.php/MiPNet14.13\\_Medium-MiR06](http://wiki.oroboros.at/index.php/MiPNet14.13_Medium-MiR06)